

Donor 5106

Genetic Testing Summary

Fairfax Cryobank recommends reviewing this genetic testing summary with your healthcare provider to determine suitability.

Last Updated: 08/14/23

Donor Reported Ancestry: African American, German, Irish, Norwegian Jewish Ancestry: No

Genetic Test*	Result	Comments/Donor's Residual Risk**
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Chromosome analysis (karyotype)	Normal male karyotype	No evidence of clinically significant chromosome abnormalities
Hemoglobin evaluation	Normal hemoglobin fractionation and MCV/MCH results	Reduced risk to be a carrier for sickle cell anemia, beta thalassemia, alpha thalassemia trait (aa/ and a-/a-) and other hemoglobinopathies
Cystic Fibrosis (CF) carrier screening	Negative by genotyping of 130 mutations in the CFTR gene	
Spinal Muscular Atrophy (SMN1)	Negative for deletions in exon 7	
Genetic Panel of 22 genes (completed in 2015)- see attached	Negative by genotyping	
Tay Sachs enzyme analysis	Negative by Hexosaminidase A enzyme analysis	

^{*}No single test can screen for all genetic disorders. A negative screening result significantly reduces, but cannot eliminate, the risk for these conditions in a pregnancy.

^{**}Donor residual risk is the chance the donor is still a carrier after testing negative.



Ordering Practice:

Practice Code: 1138 Fairfax Cryobank -

Physician:
Report Generated: 2015-08-28

5106

DOB: Gender: Male

Ethnicity: European and African

Procedure ID: 28401

Kit Barcode:

Method: Genotyping Specimen: Blood, #29853 Specimen Collection: 2015-08-20

Specimen Received: 2015-08-20 Specimen Analyzed: 2015-08-21 Partner Not Tested

SUMMARY OF RESULTS

NO MUTATIONS IDENTIFIED

5106 was not identified to carry any of the mutations tested.

All mutations analyzed were not detected, reducing but not eliminating your chance to be a carrier for the associated genetic diseases. A list of all the diseases and mutations you were screened for is included later in this report. The test does not screen for every possible genetic disease.

For disease information, please visit www.recombine.com/diseases. To speak with a Genetic Counselor, call 855.OUR.GENES.

of Male

Panel: Fairfax Cryobank Panel, Diseases Tested: 21, Mutations Tested: 382, Genes Tested: 22, Null Calls: 0

Assay performed by Reprogenetics
CLIA ID: 31 D 1054821
Lab Technician Bo Chu

Reviewed by Pere Colls, PhD, HCLD, Lab Director





Methods and Limitations

Genotyping: Genotyping is performed using the Illumina Infinium Custom HD Genotyping assay to identify mutations in >200 genes. The assay is not validated for homozygous mutations, and it is possible that individuals affected with disease may not be accurately genotyped.

Spinal Muscular Atrophy: Spinal Muscular Atrophy is tested for via an Identity-by-State shared haplotype comparison algorithm. Detection is limited to haplotypes within our library of known carriers of the most common mutation (deletion of Exon 7).

Limitations: In some cases, genetic variations other than that which is being assayed may interfere with mutation detection, resulting in false-negative or false-positive results. Additional sources of error include, but are not limited to: sample contamination, sample mixup, bone marrow transplantation, blood transfusions, and technical errors.

The test does not test for all forms of genetic disease, birth defects, and intellectual disability. All results should be interpreted in the context of family history; additional evaluation may be indicated based on a history of these conditions. Additional testing may be necessary to determine mutation phase in individuals identified to carry more than one mutation in the same gene. All mutations included within the genes assayed may not be detected, and additional testing may be appropriate for some individuals.





● High Impact ● Treatment Benefits ● X-Linked ● Moderate Impact

Diseases & Mutations Assayed

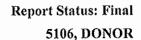
	<u> </u>		
H T X M			Mutations
• 0 0 0	Alpha Thalassemia	10	of Genotyping SEA deletion, 11.1kb deletion, c.207C>A (p.N69K), c.223G>C (p.D75G), c.2T>C (p.M1T), c.207C>G (p.N69K), c.340_351delCTCCCCGCCGAG (p.L114_E117del), c.377T>C (p.L126P), c.427T>C (p.X143Qexf32), c.*+94A>G
	Beta Thalassemia	83	d' Genotyping c.17_18delCT, c.20delA (p.E7Gfs), c.217insA (p.S73Kfs), c.223+702_444+342del620insAAGTAGA, c.230delC, c.25_26delAA, c.315+1G>A, c.315+2T>C, c.316-197C>T, c.316-146T>G, c.315+745C>G, c.316-1G>A, c.316-1G>C, c.316-2A>G, c.316-3C>A, c.316-3C>G, c.4delG (p.V2Cfs), c.51delC (p.K18Rfs), c.93-21G>A, c.92+1G>A, c.92+5G>A, c.92+5G>C, c.92+5G>T, c.92+6T>C, c.93-1G>A, c.93-1G>T, c50A>C, c.α-78g, c.α-79g, c.α-81g, c.A52T (p.K18X), c.c-137g, c.c-138t, c.c-151t, c.C118T (p.Q40X), c.G169C (p.G57R), c.G295A (p.V99M), c.G34A (p.V12l), c.G415C (p.A139P), c.G47A (p.W16X), c.G48A (p.W16X), c.t-80a, c.T2C (p.M1T), c.T75A (p.G25G), c.444+111A>G, c.g-29a, c.68_74delAAGTTGG, c.G92C (p.R31T), c.27_28insG, c.92+1G>T, c.92+1G>C, c.93-15T>G, c.93-1G>C, c.112delT, c.G113A (p.W38X), c.G114A (p.W38X), c.126delC, c.444+113A>G, c.250delG, c.225delC, c.383_385delAGG (p.Q128_A129delQAinsP), c.321_322insG (p.N109fs), c.316-1G>T, c.316-2A>C, c.316-106C>T, c.287_288insA (p.L97fs), c.271G>T (p.E91X), c.203_204delTG (p.V68Afs), c.154delC (p.P52fs), c.135delC (p.F46fs), c.92+2T>A, c.92+2T>C, c.90C>T (p.G30G), c.59A>G (p.N109), c.46delT (p.W16Gfs), c.45_46insG (p.L16fs), c.36delT (p.T13fs), c.2T>G (p.M1R), c.1A>G (p.M1V), c.c-137t, c.c-136g, c.c-142t, c.c-140t
• 0 0 0	Bloom Syndrome	24	Ø Genotyping c.2207_2212delATCTGAinsTAGATTC (p.Y736Lfs), c.2407insT, c.557_559delCAA (p.S186X), c.1284G>A (p.W428X), c.1701G>A (p.W567X), c.1933C>T (p.Q645X), c.C2528T (p.T843I), c.C2695T (p.R899X), c.G3107T (p.C1036F), c.2923delC (p.Q975K), c.3558+1G>T, c.3875-2A>G, c.2074+2T>A, c.2343_2344dupGA (p.781EfsX), c.380delC (p.127Tfs), c.3564delC (p.1188Dfs), c.4008delG (p.1336Rfs), c.C947G (p.S316X), c.2193+1_2193+9del9, c.C1642T (p.Q548X), c.3143delA (p.1048NfsX), c.356_357delTA (p.Cys120Hisfs), c.4076+1delG, c.C3281A (p.S1094X)
•000	Canavan Disease	8	of Genotyping c.433-2A>G, c.A854C (p.E285A), c.C693A (p.Y231X), c.C914A (p.A305E), c.A71G (p.E24G), c.C654A (p.C218X), c.T2C (p.M1T), c.G79A (p.G27R)



нтхм			Mutations
	Cystic Fibrosis	130	σ' Genotyping c.1029delC, 1153_1154insAT, c.1519_1521delATC (p.507dell), c.1521_1523delCTT (p.508delF), c.1545_1546delTA (p.Y515Xfs), c.1585-1G>A, c.164+12T>C, c.1680-886A>G, c.1680-1G>A, c.1766+1G>A, c.1766+1G>T, c.1766+5G>T, c.1818del84, c.1911delG, c.1923delCTCAAAACTinsA, c.1973delGAAATTCAATCCTinsAGAAA, c.2052delA (p.K684fs), c.2052insA (p.Q685fs), c.2051_2052delAAinsG (p.K6845fsX38), c.2174insA, c.261delTT, c.2657+5G>A, c.273+1G>A, c.273+3A>C, c.274+1G>A, c.2988+1G>A, c.3039delC, c.3140-26A>G, c.325delTATinsG, c.3527delC, c.3535delACCA, c.3691delT, c.3717+12191C>T, c.3744delA, c.3773_3774insT (p.11258fs), c.442delA, c.489+1G>T, c.531delT, c.579+1G>T, c.579+5G>A (IVS4+5G>A), c.803delA (p.N268fs), c.805_806delAT (p.1269fs), c.933_935delCTT (p.311delF), c.A1645C (p.S549R), c.A2128T (p.K710X), c.C1000T (p.R334W), c.C1013T (p.T338I), c.C1364A (p.A455E), c.C1477T (p.Q493X), c.C1572A (p.C524X), c.C1657T (p.Q552X), c.C1657T (p.R553X), c.C721A (p.P574H), c.C2125T (p.R709X), c.C3472T (p.R1158X), c.C3484T (p.R1162X), c.C349T (p.R117C), c.C3587G (p.Y1092X), c.C3472T (p.R1158X), c.C3448T (p.R1162X), c.C349T (p.R117C), c.G3587G (p.S1196X), c.G346A (p.R347H), c.G1040C (p.R347P), c.G1438T (p.G480C), c.G1624T (p.G542X), c.G1646A (p.S549N), c.G1643T (p.E92X), c.G3209A (p.R1070Q), c.G3266A (p.M1089X), c.G3752A (p.G353A (p.G350A (p.R117H), c.G1675A (p.G552A), c.G356A (p.G622D), c.G254A (p.G85E), c.G271A (p.G91R), c.G274T (p.E92X), c.G3209A (p.R1070Q), c.G3266A (p.M1089X), c.G3454C (p.D1152H), c.G350A (p.R117H), c.G3611A (p.W1204X), c.G3752A (p.G350A), c.T1090C (p.S364P), c.T3302A (p.M101K), c.T617G (p.1206W), c.C14T (p.P51), c.G1675A (p.S549R), c.G171A (p.W57X), c.313delA (p.1105fs), c.G328C (p.D110H), c.580-1G>7, c.G1055A (p.R352Q), c.C1075A (p.G350A (p.G178R), c.G988T (p.G330X), c.T1090C (p.S364P), c.T3302A (p.M1070Q), c.G3266A (p.W1089X), c.G386B (p.Y1080X), c.G171A (p.S549R), c.G171A (p.W57X), c.313delA (p.1105fs), c.G328C (p.D110H), c.580-1G>7, c.G1055A (p.R352Q), c.C1075A (p.G358K), c.C1079A (p.T360K), c.T1647G (p.S54
• 0 0 0	Familial Dysautonomia	4	of Genotyping c.2204+6T>C, c.C2741T (p.P914L), c.G2087C (p.R696P), c.C2128T (p.Q710X)
• 0 0 0	Familial Hyperinsulinism: Type 1: ABCC8 Related	10	of Genotyping c.3989-9G>A, c.4159_4161delTTC (p.1387delF), c.C4258T (p.R1420C), c.C4477T (p.R1493W), c.G2147T (p.G716V), c.G4055C (p.R1352P), c.T560A (p.V187D), c.4516G>A (p.E1506K), c.C2506T (p.Q836X), c.579+2T>A
••00	Fanconi Anemia: Type C	8	d Genotyping c.456+4A>T, c.67delG, c.C37T (p.Q13X), c.C553T (p.R185X), c.T1661C (p.L554P), c.C1642T (p.R548X), c.G66A (p.W22X), c.G65A (p.W22X)
	Gaucher Disease	6	of Genotyping c.84_85insG, c.A1226G (p.N409S), c.A1343T (p.D448V), c.C1504T (p.R502C), c.G1297T (p.V433L), c.G1604A (p.R535H)
	Glycogen Storage Disease: Type IA	13	of Genotyping c.376_377insTA, c.79delC, c.979_981delTTC (p.327delF), c.C1039T (p.Q347X), c.C247T (p.R83C), c.C724T (p.Q242X), c.G248A (p.R83H), c.G562C (p.G188R), c.G648T, c.G809T (p.G270V), c.A113T (p.D38V), c.975delG (p.L326fs), c.724delC
•000	Joubert Syndrome	1	o [®] Genotyping c.G35T (p.R12L)
	Maple Syrup Urine Disease: Type 1B	6	of Genotyping c.G1114T (p.E372X), c.G548C (p.R183P), c.G832A (p.G278S), c.C970T (p.R324X), c.G487T (p.E163X), c.C853T (p.R285X)



н т х м			Mutations
	Maple Syrup Urine Disease: Type 3	8	o ^a Genotyping c.104_105insA, c.G685T (p.G229C), c.A214G (p.K72E), c.A1081G (p.M361V), c.G1123A (p.E375K), c.T1178C (p.1393T), c.C1463T (p.P488L), c.A1483G (p.R495G)
• 0 0 0	Mucolipidosis: Type IV	4	o [®] Genotyping c.406-2A>G, c.G1084T (p.D362Y), c.C304T (p.R102X), c.244delC (p.L82fsX)
•000	Nemaline Myopathy: NEB Related	1	of Genotyping c.7434_7536del2502bp
•000	Niemann-Pick Disease: Type A	6	o ^a Genotyping c.996delC, c.G1493T (p.R498L), c.T911C (p.L304P), c.C1267T (p.H423Y), c.G1734C (p.K578N), c.1493G>A (p.R498H)
• 0 0 0	Spinal Muscular Atrophy: SMN1 Linked	19	of Genotyping DEL EXON 7, c.22_23insA, c.43C>T (p.Q15X), c.91_92insT, c.305G>A (p.W102X), c.400G>A (p.E134K), c.439_443delGAAGT, c.558delA, c.585_586insT, c.683T>A (p.L228X), c.734C>T (p.P245L), c.768_778dupTGCTGATGCTT, c.815A>G (p.Y272C), c.821C>T (p.T274I), c.823G>A (p.G275S), c.834+2T>G, c.835-18_835-12delCCTTTAT, c.835G>T, c.836G>T
• 0 0 0	Tay-Sachs Disease	30	of Genotyping c.1073+1G>A, c.1277_1278insTATC, c.1421+1G>C, c.805+1G>A, c.C532T (p.R178C), c.G533A (p.R178H), c.G805A (p.G269S), c.C1510T (p.R504C), c.G1496A (p.R499H), c.G509A (p.R170Q), c.A1003T (p.1335F), c.910_912delTTC (p.305delF), c.G749A (p.G250D), c.T632C (p.F211S), c.C629T (p.S210F), c.613delC, c.A611G (p.H204R), c.G598A (p.V200M), c.A590C (p.K197T), c.571-1G>T, c.C540G (p.Y180X), c.T538C (p.Y180H), c.G533T (p.R178L), c.C508T (p.R170W), c.C409T (p.R137X), c.T380G (p.L127R), c.346+1G>C, c.T116G (p.L39R), c.G78A (p.W26X), c.A1G (p.M1V)
•000	Usher Syndrome: Type 1F	6	of Genotyping c.C733T (p.R245X), c.2067C>A (p.Y684X), c.C7T (p.R3X), c.C1942T (p.R648X), c.2800C>T (p.R934X), c.4272delA (p.L1425fs)
• 0 0 0	Usher Syndrome: Type 3	4	o [®] Genotyping c.T144G (p.N48K), c.T359A (p.M120K), c.300T>G (p.Y176X), c.C634T (p.Q212X)
•000	Walker-Warburg Syndrome	1	♂ Genotyping c.1167insA (p.F390fs)



Lab:CH



Patient Information	Specimen Information	Client Information
5106, DONOR	Specimen: Requisition:	Client #: 9595 Not Provided
DOB: AGE:	Lab Ref#:	
Gender: M Phone: NG	Collected: 08/20/2015 / 00:00 EDT	
Patient ID: NG	Received: 08/21/2015 / 03:10 EDT	
Patient ID: NG	Reported: 09/01/2015 / 15:06 EDT	

Cytogenetics Report

Chromosome Analysis, Blood - 14596

CB-15-014957

Specimen Source

Case Number

Peripheral Blood

Clinical History

Donor

Metaphases Counted

201101

metaphases counted

20

Metaphases Analyzed

5

Metaphases Karyotyped

.

Banding Level

>=550

Karyotype

46,XY

Interpretation and Comments

NORMAL MALE karyotype

Within the limits of standard cytogenetic methodologies, the chromosomes had normal G-banding patterns without apparent structural abnormality or rearrangement.

This test does not address genetic disorders that cannot be detected by standard cytogenetic methods, or rare events such as low level mosaicism or very subtle rearrangements.

Signature

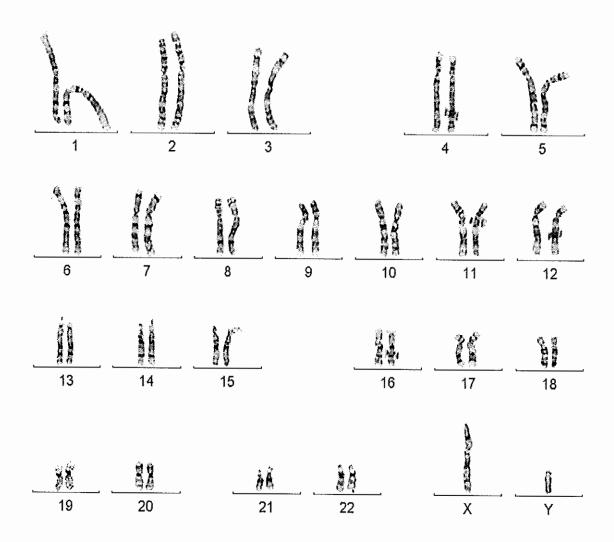
Electronic Signature on File

Nicole C. Christacos, Ph.D., FACMG

Technical Director, Cytogenetics, 703-802-7156



Patient Information	Specimen Information	Client Information
5106, DONOR DOB: AGE: Gender: M Patient ID: NG	Specimen: Collected: 08/20/2015 / 00:00 EDT Received: 08/21/2015 / 03:10 EDT Reported: 09/01/2015 / 15:06 EDT	Client #: 9595 Not Provided



PERFORMING SITE:

CH QUEST DIAGNOSTICSANICHOLS CHANTILLY, 14225 NEWBROOK DRIVE, CHANTILLY, VA 20151-0841 Laboratory Director. KENNETH SISCO, MD PHD, CLIA: 49D0221801

This is supplemental to your standard report.

CLIENT SERVICES: 866-677-0742 (Opt#1)

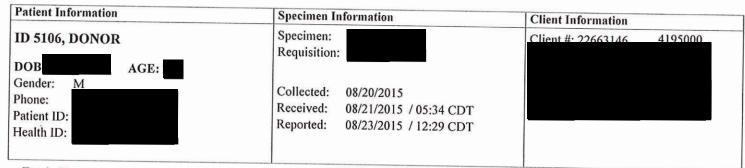
SPECIMEN:





Report Status: Final

ID 5106, DONOR



Test Name HEMOGLOBINOPATHY EVALUATION	In Range	Out Of Range	Reference Range	Lab
RED BLOOD CELL COUNT HEMOGLOBIN HEMATOCRIT MCV MCH RDW HEMOGLOBIN A	4.74 14.9 45.0 94.9 31.5 13.4		4.20-5.80 Million/uL 13.2-17.1 g/dL 38.5-50.0 % 80.0-100.0 fL 27.0-33.0 pg 11.0-15.0 %	СВ
HEMOGLOBIN F HEMOGLOBIN A2 (QUANT) INTERPRETATION	97.5 <1.0 2.5		>96.0 % <2.0 % 1.8-3.5 %	СВ

Normal phenotype.

Normal hemoglobin distribution, no HgS, HgC or other abnormal hemoglobin observed.



PERFORMING SITE:

CB QUEST DIAGNOSTICS WOOD DALE, 1355 MITTEL BOULEVARD, WOOD DALE, IL 60191-1024 Laboratory Director: ANTHONY V. THOMAS, MD, CLIA: 14D0417052



Tay-Sachs Enzyme Analysis

Patient Name: 5106, Donor

Referring Physician: Specimen #:

Client #:

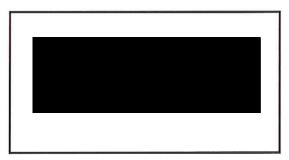
Patient ID:

DOB: SSN: ***-**- Date Collected: 08/20/2015 Date Received: 08/21/2015

Lab ID: 5106-150820

Hospital ID:

Specimen Type: White Blood Cells



ENTERED Co-9/9/15

RESULTS:

Hexosaminidase Activity: 1322 nmol/mg protein

Hexosaminidase Percent A: 54.5

Plasma/Serum

WBC

Expected Non-Carrier Range:

Hex A ≥54%

≥54%

Expected Carrier Range:

Hex A 20 - 49%

20 - 49%

INTERPRETATION: NON CARRIER

This result is within the non-carrier range for Tay-Sachs disease. Less than 0.1% of patients having non-carrier levels of Hexosaminidase-A activity are Tay-Sachs carriers.

NOTE: Maximum sensitivity and specificity for Tay-Sachs disease carrier testing are achieved by using enzymology and DNA mutation analysis together.

anterel Warenbery, PHO, MOCC

Integrated Genetics is a business unit of Esoterix Genetic Laboratories, LLC, a wholly-owned subsidiary of Laboratory Corporation of America Holdings.

Under the direction of:

Stanford Marenberg, Ph.D.

Date: 08/28/2015

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Testing Performed At Esoterix Genetic Laboratories, LLC 2000 Vivigen Way Santa Fe, NM 87505 Philip Wyatt, MD, PhD, Laboratory Director 1-800-848-4436